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### A novel preparation of D-fructopyranose 5-sulphate

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(Received August 3rd, 1982; accepted for publication, September 10th, 1982)

We have described<sup>1</sup> the synthesis and characterisation of a series of galactopyranose and glucopyranose sulphates, and their use in an investigation<sup>2</sup> of the specificity of sulphatase A. For the completion of this work, D-glucofuranose 5-sulphate was sought. The preparation<sup>3</sup> of D-glucofuranose 5-phosphate from 3-*O*-acetyl-1,2-*O*-isopropylidene-6-*O*-triphenylmethyl- $\alpha$ -D-glucofuranose suggested a route to D-glucofuranose 5-sulphate.

The reaction of 3,6-di-*O*-acetyl-1,2-*O*-isopropylidene- $\alpha$ -D-glucofuranose with pyridine-sulphur trioxide followed by deacetylation gave the expected 1,2-*O*-isopropylidene- $\alpha$ -D-glucofuranose 5-(barium sulphate) (**1**), which was characterised by <sup>13</sup>C-n.m.r. spectroscopy (Table I). As with other monosaccharide sulphates<sup>1</sup>, there was a large downfield shift (7.33 p.p.m.) in the signal for C-5, which carries the sulphate group, and smaller upfield shifts (1.61 and 2.57 p.p.m., respectively) in the signals for C-4 and C-6.

Autohydrolysis of the free-acid form of **1** removed the isopropylidene group and gave a sulphate ester of a reducing sugar, but the p.m.r. spectrum showed that it was not D-glucofuranose 5-sulphate because there was no signal for an anomeric proton. Also, the <sup>13</sup>C-n.m.r. spectrum was not consistent with a glucofuranose derivative. Proton-coupled, <sup>13</sup>C-n.m.r. spectroscopy showed that the compound contained one quaternary, three secondary, and two primary carbon atoms, and the spectrum was consistent with D-fructopyranose 5-(potassium sulphate). There was a large downfield shift (8.32 p.p.m.) in the signal for C-5, and smaller upfield shifts (1.46 and 2.47 p.p.m., respectively) in the signals for C-4 and C-6. As shown in Table I, the deuterium-induced, differential isotope shifts (d.i.s. shifts)<sup>4</sup> confirmed these assignments. Only for C-2 was there a significant discrepancy between the observed and calculated d.i.s. values, and a similar low value (0.12) has been observed<sup>4</sup> for C-2 in  $\beta$ -D-fructopyranose.

D-Fructose 5-sulphate was a substrate for sulphatase A: with  $K_m$  and  $V$  values of 48 mM and 13  $\mu\text{mol} \cdot \text{mg}^{-1} \cdot \text{min}^{-1}$ , respectively: these values are comparable to

TABLE I

<sup>13</sup>C-N.M.R. DATA FOR 1,2-O-ISOPROPYLDENE-D-GLUCOSE, D-FRUCTOSE, AND THEIR 5-SULPHATES

	Chemical and d.l.s. shifts (p.p.m.) <sup>a</sup>						
	C-1	C-2	C-3	C-4	C-5	C-6	C-H <sub>3</sub>
1,2-O-Isopropylidene-D-glucofuranose	105.43	85.14	74.33	80.51	69.16	64.23	113.42 26.31 25.88
1,2-O-Isopropylidene-D-glucopyranose	105.42	84.93	74.27	78.90	76.69	61.66	113.57 26.36 25.92
5-(barium sulphate)	64.91	98.89	68.57	70.68	70.16	64.24	— — —
β-D-Fructopyranose	64.64	98.90	68.50	69.22	78.48	61.77	— — —
β-D-Fructopyranose 5-(potassium sulphate)	0.19 (0.18) <sup>b</sup>	0.11 (0.17)	0.22 (0.20)	0.16 (0.17)	0.00 (0.03)	0.00 (0.00)	— — —

<sup>a</sup><sup>13</sup>C-Chemical shifts were measured with respect to internal 1,4-dioxane and converted to the Mc/Sr scale by using the relationship  $\delta_{Mc/Sr} = \delta_{dioxane} - 67.40$ . <sup>b</sup>Values are for the observed and (in brackets) calculated d.l.s. values<sup>1</sup> for D-fructose 5-sulphate.

those for D-glucose 3-sulphate<sup>2</sup>. Moreover, the liberated sugar had the chromatographic properties of fructose (see Experimental).

Unlike gluco- and galacto-pyranose sulphates<sup>1</sup>, the fructopyranose 5-sulphate was precipitated by ethanol as a single anomer, as shown by <sup>13</sup>C-n.m.r. spectroscopy. D-Fructopyranose exists<sup>4</sup> in solution mainly as the  $\beta$  anomer, and this is likely also for the 5-sulphate which had an  $[\alpha]_D$  value (water) of  $-86^\circ$ .

The conversion of a 5-substituted 1,2-*O*-isopropylidene- $\alpha$ -D-glucofuranose into a 5-substituted  $\beta$ -D-fructopyranose apparently has not been described hitherto and presumably involves a Lobry de Bruyn-Alberda van Ekenstein transformation<sup>5</sup> of D-glucofuranose 5-sulphate. The slow transformation of glucose into fructose in acid solution is long known<sup>6</sup>, but the conversion noted here, as judged by the disappearance of the n.m.r. signal for the anomeric proton, was essentially complete in 30 min at  $70^\circ$ . The reaction is also unusual in that the ketose is the main product, whereas, in most such transformations, a complex mixture of sugars is obtained<sup>5</sup>. If the transformation is a general one, it could be useful for the synthesis of 5-substituted fructopyranoses, although other routes<sup>7</sup> are available.

The above results cast doubt on the structure of the presumed D-glucofuranose 5-phosphate<sup>3</sup>, but it should be noted that Fitzgerald<sup>8</sup> suggested a specific role for the sulphate group in the Tris-catalysed isomerisation of glucose 6-sulphate to fructose 6-sulphate.

#### EXPERIMENTAL

*General methods.* — The general chemical and spectroscopic methods were described in the previous paper<sup>1</sup>.

*1,2-O-Isopropylidene- $\alpha$ -D-glucofuranose 5-(barium sulphate).* — 3,6-Di-*O*-acetyl-1,2-*O*-isopropylidene- $\alpha$ -D-glucofuranose<sup>9</sup> (7.3 g, 24 mmol) was treated overnight with pyridine-sulphur trioxide (5.8 g, 36 mmol) in pyridine (100 mL) at room temperature and the product (13 g) was isolated by the usual methods<sup>1</sup>. This was deacetylated with methanolic barium methoxide<sup>1</sup> to give the crude, title compound (10 g). P.m.r. data (D<sub>2</sub>O):  $\delta$  5.83 (d,  $J_{1,2}$  3.66 Hz, H-1).

*$\beta$ -D-Fructopyranose 5-(potassium sulphate).* — The foregoing compound (two lots of 4.0 and 6.0 g) was converted into the free acid by passage through a column of Dowex-50(H<sup>+</sup>) resin and the acidic eluate ( $\sim 100$  mL for each lot) was kept for 40 min at  $80^\circ$ . Removal of the isopropylidene group was confirmed by p.m.r. spectroscopy, and the barium salt of the sulphate ester was obtained in the usual manner<sup>1</sup>.

This crude barium salt (10 g) was converted into the free acid, as described above, and thence<sup>10</sup> into the brucine salt. Three recrystallisations from aqueous acetone<sup>10</sup> gave a product (3.6 g), m.p.  $176-178^\circ$  (dec.),  $[\alpha]_D -65^\circ$  (water).

*Anal.* Calc. for C<sub>6</sub>H<sub>11</sub>O<sub>9</sub>S · C<sub>23</sub>H<sub>27</sub>N<sub>2</sub>O<sub>4</sub>: C, 53.2; H, 5.85; N, 4.28; S, 4.90. Found: C, 52.9; H, 4.57; N, 4.17; S, 4.57.

The pure brucine salt (2.3 g) was converted<sup>10</sup> into the potassium salt, which was precipitated from aqueous solution with ethanol<sup>1</sup> to give the title compound

(0.9 g),  $[\alpha]_D -86^\circ$  (water). The tenaciously retained ethanol of solvation was detectable by  $^{13}\text{C}$ -n.m.r. spectroscopy, and there was no n.m.r. signal ( $\text{D}_2\text{O}$ ) for an anomeric proton.

*Anal.* Calc. for  $\text{C}_6\text{H}_{11}\text{KO}_5\text{S}$ : C, 24.2; H, 3.72; S, 10.8. Found: C, 23.9; H, 4.02; S, 9.28.

*Enzymic hydrolysis of D-fructopyranose 5-sulphate.* — A solution of D-fructose 5-(potassium sulphate) (25  $\mu\text{mol}$ ) in 0.1M pyridine-acetic acid buffer (pH 5.6, 0.25 mL) was treated with 250  $\mu\text{g}$  of sulphatase A for 4 days at room temperature, when analysis<sup>2</sup> revealed  $\sim 50\%$  hydrolysis. Dilution (10-fold) of a sample of the hydrolysate with water followed by t.l.c.<sup>11</sup> [phosphate-impregnated Silica gel 60 (Merck), acetone-2-propanol-0.1M lactic acid (2:2:1), detection with orcinol<sup>12</sup> and 1-naphthol-phosphoric acid<sup>13</sup>] revealed fructose,  $R_f$  0.20 (*cf.* 0.26 for glucose)

## REFERENCES

- 1 P. J. ARCHBALD, M. D. FENN, AND A. B. ROY, *Carbohydr. Res.*, **93** (1981) 177-190.
- 2 A. B. ROY AND J. TURNER, *Biochim. Biophys. Acta*, **704** (1982) 366-373.
- 3 K. JOSEPHSON AND S. PROFFE, *Biochem. Z.*, **258** (1933) 147-153.
- 4 P. E. PFEFFER, K. M. VALENTINE, AND F. W. PARISH, *J. Am. Chem. Soc.*, **101** (1979) 1265-1274.
- 5 J. C. SPECK, JR., *Adv. Carbohydr. Chem.*, **13** (1958) 63-103.
- 6 H. OST, *Z. Angew. Chem.*, **18** (1905) 1170-1174.
- 7 K. HEYNS AND J. HEUKSHOVEN, *Justus Liebigs Ann. Chem.*, (1976) 269-283.
- 8 J. W. FITZGERALD, *Can. J. Biochem.*, **53** (1975) 906-910.
- 9 K. FREUDENBERG AND K. VON OERTZEN, *Justus Liebigs Ann. Chem.*, **574** (1951) 37-53.
- 10 K. B. GUISELEY AND P. M. RUOFF, *J. Org. Chem.*, **26** (1961) 1248-1254.
- 11 S. A. HANSEN, *J. Chromatogr.*, **107** (1975) 224-226.
- 12 L. HOUGH, J. K. N. JONES, AND W. H. WADMAN, *J. Chem. Soc.*, (1950) 1702-1706.
- 13 N. ALBON AND D. GROSS, *Analyst*, **75** (1950) 454-457.